

THE USE OF BIOLOGICAL APPROACHES IN THE TREATMENT OF SHOULDER PATHOLOGY

A Critical Analysis Review

Jonas Pogorzelski, MD, MHBA Jonathan A. Godin, MD, MBA Erik M. Fritz, MD Mark E. Cinque, MS Jorge Chahla, MD Johnny Huard, PhD Peter J. Millett, MD, MSc

Investigation performed at the Steadman Philippon Research Institute, Vail, Colorado

Abstract

- » The major pathological conditions affecting the shoulder that are treated with use of biological applications include focal cartilage lesions and rotator cuff tears. Biological modalities that previously have been used or investigated include platelet-rich plasma (PRP), growth factors, progenitor cells, bone-marrow stimulation, autologous chondrocyte implantation (ACI), matrix-induced ACI (MACI), and biological scaffolds.
- » Marrow-stimulating procedures have been reported to yield positive results when used for the treatment of focal cartilage lesions of the glenoid or humeral head. Limited data are available on the use of PRP, ACI, and MACI for the treatment of chondral lesions involving the shoulder, and therefore no conclusions can be drawn regarding the efficacy of these modalities.
- » Preclinical and in vitro studies have demonstrated that modulation of growth factors may be helpful for rotator cuff tear healing; however, the optimal modulation and delivery vehicle remain to be elucidated. PRP has received much research attention; however, most studies have been hindered by study setup and conflicting results. Therefore, the use of PRP to enhance rotator cuff healing remains controversial. Progenitor cells have shown positive results in a small number of preclinical and clinical studies, but further research is needed before conclusions can be drawn.
- » In summary, basic-science studies investigating biological factors to enhance healing in the shoulder have shown potential. However, clinical data are still limited, contradictory, and controversial. Additional research is needed. Most importantly, robust, consistent, well-powered clinical trials are necessary to definitively determine which methods improve clinical outcomes.

houlder pain is the second most common musculoskeletal complaint encountered in primary care offices, occurring in about 51% of patients¹. Over the last few years, major advances have been made in both the operative and nonoperative treatment of shoulder pain and pathological conditions.

However, because of increasing patient age and the rising number of active patients, there is a demand for improved treatment of all shoulder abnormalities. Therefore, the use of orthobiologics in shoulder surgery has expanded rapidly over the past decade. Some of the commonly used biologics in shoulder surgery include

COPYRIGHT © 2017 BY THE JOURNAL OF BONE AND JOINT SURGERY, INCORPORATED **Disclosure:** No funding was received during the preparation or execution of this manuscript. On the **Disclosure of Potential Conflicts of Interest** forms, *which are provided with the online version of the article*, one or more of the authors checked "yes" to indicate that the author had a relevant financial relationship in the biomedical arena outside the submitted work (https://links.lww.com/JBJSREV/A265).



progenitor cells, growth factors, plateletrich plasma (PRP), and biological matrices. The potential advantages of biological augmentation of traditional shoulder surgical techniques include minimal invasiveness, improved healing capacity, and more-rapid recovery. Conversely, the use of biologics is currently expensive, and the evidence of long-term effectiveness is limited. Furthermore, the body of literature on the use and efficacy of biologics in shoulder surgery is heterogeneous with regard to indications, therapies, processing methods, and inoculation. Although some studies have demonstrated encouraging results following either isolated treatment with biologics or biologic-augmented surgery, other studies have failed to demonstrate substantial benefit. Given the heterogeneity and paucity of critical analyses within the literature, the objective of the present review is to evaluate the value of orthobiologics in the treatment of shoulder pathologies. More precisely, we aim to perform a balanced evaluation of biologic augmentation of existing modalities for the treatment of focal chondral defects, osteoarthritis, and rotator cuff tears.

Focal Chondral Defects and Glenohumeral Osteoarthritis

A trial of conservative therapy consisting of physical therapy and nonsteroidal anti-inflammatory medications is typically seen as the primary treatment when a symptomatic cartilage defect is identified in the shoulder². If symptoms persist, biologic-based treatments such as marrow-stimulation procedures, bone marrow aspirate concentrate (BMAC), PRP, and cell-based therapies such as autologous chondrocyte implantation (ACI) or matrix-induced ACI (MACI) can be used.

Microfracture

The microfracture procedure has been widely used in several joints because of its ease of performance, low cost, and overall positive reported outcomes when used for the treatment of small,

contained cartilage lesions. The aim of this technique is to perforate the subchondral bone to promote the release of progenitor cells and multiple growth factors to the chondral defect. Under favorable mechanical-stress characteristics (stress-free joint movement with varying pressure, tensile, and shearing forces), these progenitor cells will differentiate into fibrochondrocytes, ultimately creating a fibrocartilage layer. However, the higher content of type-I collagen makes this repair tissue one of lesser biomechanical quality than hyaline cartilage, as previously reported in animal experiments³. Patient selection is most certainly key for successful outcomes. However, clear indications are not available in the shoulder literature; therefore, on the basis of an algorithm that has been extrapolated from the knee literature, microfracture is considered to be suitable for the treatment of small, circumscribed cartilage defects (maximum, 4 cm²) with intact subchondral bone in young, active patients⁴⁻⁷.

In general, better results have been reported for small unipolar defects on either the glenoid or humeral side, whereas the worst results have been seen in association with bipolar glenohumeral lesions^{2,3,8}. Millett et al., in a study of 31 shoulders, reported significant (p < 0.05) reduction in pain and improvement in function according to the American Shoulder and Elbow Surgeons (ASES) score⁹ at a mean of 4 years after the use of microfracture for the treatment of full-thickness articular cartilage injuries. Only 19% of the patients in that cohort were considered to have had a failure requiring additional surgery². Likewise, Frank and colleagues, in a series of 16 patients (17 shoulders), reported significant improvement in shoulder function as demonstrated by an increase in the Simple Shoulder Test (SST) and ASES scores (p < 0.01 for both) and significant pain reduction (p < 0.01) at >2 years postoperatively¹⁰. Outcomes did not significantly vary with respect to sex or age. Of note, 93%

of the patients stated that they would have the surgery again. Last, Siebold and colleagues 11 reported significant improvements in pain and functional scores (p = 0.0053 and p = 0.018, respectively) in patients managed with microfracture combined with a periosteal flap. Poor prognostic predictors included prior surgery and, potentially, lesion size, although the results were not significant.

Platelet-Rich Plasma (PRP)

PRP has been widely utilized for the treatment of various musculoskeletal conditions or as an augmentation tool on the basis of basic-science and emerging clinical studies 12,13. The biological reasoning behind the clinical use of PRP includes local delivery of growth factors, modification of the inflammatory response, increased hemostasis, and positive effects on cell proliferation and differentiation 14-17. Although symptomatic relief has been reported following the use of PRP for the treatment of early-stage knee osteoarthritis¹⁴, we are not aware of any studies that have evaluated the efficacy of PRP in the shoulder. However, autologous platelet-poor plasma and platelet gel have been shown to reduce pain scores and to yield significantly better functional outcomes (internal rotation, p < 0.05) following total shoulder arthroplasty¹⁸. In the study by Lo et al. 19, 55 patients with glenohumeral arthritis were managed with a biologically based resurfacing arthroplasty with use of acellular human dermal allograft in combination with PRP. After an average duration of follow-up of 60 months, the average ASES score was 76 ± 22 , the average Western Ontario Osteoarthritis of the Shoulder (WOOS) index was 76% ± 22%, and the average visual analog scale (VAS) score for pain was 2.4 ± 2.6 , with an 81% rate of patient satisfaction. The average joint space increased from 1 ± 1 mm preoperatively to 2 ± 1 mm postoperatively. Fewer than 10% of the patients underwent revision to total shoulder arthroplasty.



Autologous Chondrocyte Implantation (ACI) and Matrix-Induced ACI (MACI)

The principle of these techniques is to culture autologous cells and then to implant these cells into the chondral defect²⁰. These procedures can be performed with or without a 3-dimensional biocompatible scaffold. ACI and MACI are staged procedures in which an initial arthroscopic harvest is performed to obtain chondrocytes for culture. The chondrocytes are then expanded in culture to obtain 15 to 20 million cells. This expansion takes approximately 1 month, and the cells are then implanted into the chondral defect. There is a paucity of data in the orthopaedic literature on the use of ACI for the treatment of chondral defects within the shoulder. We are aware of only a single case in which ACI has been used for the treatment of a shoulder defect; in that report, a 16year-old male athlete with a full-thickness lesion (3.3 \times 1.5 cm) of the humeral head reported functional improvements at 12 months²¹.

For the MACI procedure, a few days prior to implantation, a biodegradable scaffold is seeded with the expanded chondrocytes, which can then synthesize extracellular matrix components²¹. We are aware of only 1 small report on the use of this procedure in the shoulder²². In that study, 4 young adults were managed with ACI for the treatment of symptomatic, isolated, large-diameter lesions of the cartilage involving the humerus (3 patients; defect size, 6 cm²) or glenoid (1 patient; defect size, 2 cm²). After a mean duration of follow-up of >3 years, the mean pain and functional scores were considered satisfactory (VAS score for pain, 0.3 of 10; Constant score, $83.3 \pm$ 9.9; and ASES index, 95.3 ± 8.1). Moreover, the coverage of the defect on magnetic resonance imaging (MRI) also was deemed satisfactory, with signs of fibrocartilaginous repair tissue.

Rotator Cuff Tears

A number of novel approaches have been described to enhance the biological healing of the rotator cuff repair site, to improve the regeneration of the native cuff insertion site, and to inhibit the formation of scar tissue^{23,24}. These approaches include tissue engineering, cell therapy, and growth factors.

Marrow-Venting Procedures

Bleeding bone surfaces, such as those brought about by venting of the greater tuberosity or acromioplasty, can enable the release of growth factors, which can in turn lead to improved cuff healing^{22,25}. These factors, including platelet-derived growth factor (PDGF), basic fibroblast growth factor (bFGF), and vascular endothelial growth factor (VEGF), induce inflammation and angiogenesis. Meanwhile, other growth factors improve matrix synthesis, cellular proliferation, and cellular differentiation (transforming growth factor-beta [TGF-β]), promote osseous incorporation of tendon (bone morphogenetic proteins [BMPs]), and remodel the extracellular matrix (matrix metalloproteinases [MMPs])²⁶. Gotoh et al., in a study of 24 patients with or without a retear following rotator cuff repair, found increased levels of MMP-3 and tissue inhibitor of MMP (TIMP)-1 in the retear group and concluded that these findings may suggest a potential approach for targeted drug therapy following rotator cuff repair²⁶. While there have been no studies investigating rotator cuff healing following treatment with recombinant growth factors in humans, the results of preclinical studies have indicated that the delivery or modulation of these factors may augment rotator cuff healing^{27,28}. Bedi et al., on the basis of their findings in a rat model, reported that MMP-13 activity modulation with doxycycline following rotator cuff repair may offer a novel biological approach to improve tendon-tobone healing²⁹. However, the clinical translation of these in vitro and animal studies remains a challenge.

Microfracture

Microfracture of the greater tuberosity, immediately lateral to the rotator cuff repair site, results in extravasation of mesenchymal stromal cells (MSCs), platelets, and growth factors. However, several studies have shown mixed results. Osti et al., in a randomized controlled trial of 57 patients who underwent arthroscopic rotator cuff repair, demonstrated that microfracture of the rotator cuff footprint resulted in reduced shortterm pain but did not result in significantly different long-term clinical or radiographic outcomes³⁰. Milano et al., in prospective randomized study of 80 patients who underwent arthroscopic rotator cuff repair with or without greater tuberosity microfracture, found no significant difference between the groups on MRI evaluation; however, patients receiving microfracture for large tears demonstrated significantly improved healing rates $(p = 0.040)^{31}$.

Platelet-Rich Plasma (PRP)

The use of PRP as a biological adjuvant to enhance the healing of rotator cuff tendons has recently increased in popularity, which in turn has led to a number of published studies. PRP is a term that is used to describe preparations of whole blood enriched with platelets that, once activated, release a host of growth factors that may contribute to tissue repair. Several proteoglycans (decorin, aggrecan, and biglycan) have been shown to stimulate the production of key extracellular matrix proteins and to increase proliferation of rotator cuffderived tenocytes³². Moreover, PRP inhibits the inflammatory effects of interleukin-1B (IL-1B), which contributes to rotator cuff tendon degeneration, and heightens levels of TGF-B, which increases rotator cuff tendon repair strength^{33,34}. However, these in vitro results have not translated into clinical effects; controversial results have been reported following clinical trials involving the use of different PRP formulations to enhance rotator cuff repairs^{32,35-43}.

Multiple studies have documented significantly improved results in association with the use of PRP augmentation. Pandey et al. evaluated 102 patients who underwent rotator cuff repair with or without moderately concentrated PRP

and were followed for a minimum of 2 years 44. At 24 months, the PRP group demonstrated a significantly lower retear rate than the control group (3.8% compared with 20%; p = 0.01). The difference in retear rates, however, was significant only for large tears (p = 0.03). Doppler ultrasound demonstrated that the PRP group had significantly (p < 0.05) increased vascularity of the repair site at 3 months postoperatively and in the peribursal tissue until 12 months⁴⁴. In addition, the PRP group demonstrated significantly improved University of California-Los Angeles (UCLA) scores at 12 months postoperatively (p < 0.05) and significantly improved Constant scores at 24 months postoperatively (p < 0.05). Jo et al., in a randomized controlled trial, reported a significantly lower retear rate among patients who received PRP than among those who did not (3.0% compared with 20.0%; p =0.043)⁴⁵. Additionally, the supraspinatus cross-sectional area was significantly larger in the PRP group (p = 0.014); this finding led the authors to conclude that the quality of postoperative tendon healing is increased by the use of PRP. Of note, the speed of healing and the functional outcomes were equivalent between the 2 groups. Holtby et al., in a retrospective study of 82 patients who were followed for 6 months after the repair of small to medium-sized rotator cuff tears, reported transient improvements in perioperative pain control when comparing the PRP group with the control group 46. Moreover, neither patient-oriented outcome measures nor structural integrity of the repair significantly differed between groups.

The varying protocols and conflicting results of studies investigating PRP in the setting of rotator cuff repair have led to numerous meta-analyses to further evaluate the data¹². Warth et al., in an analysis of 11 Level-I and II studies, found that clinical outcomes did not differ between patients who received PRP and controls¹³. The authors reported that placing the PRP at the tendon-bone interface rather than over the surface of the repaired tendon was associated with an overall increase in the

Constant score. Additionally, a subgroup analysis of patients who underwent double-row repair for the treatment of large (>3-cm) rotator cuff tears showed significantly lower retear rates in association with PRP use (p = 0.046). Vavken et al. performed a costeffectiveness analysis and meta-analysis of 13 published studies on the use of PRP for the repair of small and mediumsized rotator cuff tears. In contrast to Warth et al. 13, the authors found reduced retear rates following the arthroscopic repair of small and medium-sized rotator cuff tears when patients who had been managed with PRP were compared with controls⁴⁷. However, for large and massive tears, there was no decrease in the retear rate in association with PRP. Cost analysis indicated that, with the current cost, the use of PRP was not costeffective. Chahal et al., in a systematic review, also noted a reduction in retear rates for small and medium-sized rotator cuff tears 48. Most recently, Saltzman et al., in a meta-analysis of available review studies, found that the use of PRP for rotator cuff repair did not result in significantly lower overall retear rates or improved clinical outcome scores⁴⁹. Subgroup analysis showed evidence of improved outcomes in association with solid as compared with liquid PRP matrix, small or medium-sized tears as compared with large or massive tears, PRP application at the tendon-bone interface as compared with over the tendon, and in the setting of double-row as compared with single-row repair.

Because of inconsistent clinical results, the use of PRP to improve postoperative rotator cuff healing continues to be an area of debate. Additional randomized clinical trials and basic-science studies are needed to determine the optimal formulation of PRP to improve physiological healing.

Progenitor Cells

The use of multipotent MSCs has become an area of growing interest. Specifically, adipose-derived stem cells (ADSCs) and bone marrow-derived stem cells (BMSCs) can proliferate and

differentiate into multiple musculoskeletal tissues, such as tendon, ligament, cartilage, bone, and fat. Interestingly, recent articles have demonstrated the presence of stem cells obtained from bursal tissue. Of note, when treated with BMP-12, these cells expressed markers of tenocytes; therefore, BMP-12 potentially could be an important target in the treatment of rotator cuff degeneration ^{50,51}.

While preclinical studies have demonstrated promising results⁵²⁻⁵⁴, there is still a paucity of clinical studies. Only 2 clinical studies have shown the efficacy of BMSC injections in the shoulder^{55,56}. Ellera Gomes et al. reported on 14 patients in whom fullthickness rotator cuff tears were repaired with transosseous sutures with use of a mini-open approach and subsequently augmented with 10 mL of BMSC concentrate, which was injected into the repaired tendon edges⁵⁵. After 12 months of follow-up, MRI examinations demonstrated intact tendons in all patients. Additionally, Hernigou et al. evaluated the 10-year results for 90 patients who underwent single-row rotator cuff repair with (45 patients) or without (45 patients) augmentation with concentrated BMSCs⁵⁶. Intact tendon was found in 87% of the patients in the BMSC group, compared with only 44% of those in the control group.

Adipose tissue represents an abundant and reproducible progenitor cell source as ADSCs can be easily harvested from it. We are aware of only 1 study, involving a rabbit model, which has verified the positive rotator-cuff healing effects of ADSCs⁵⁴. The authors demonstrated that muscle function and tendon integrity were improved following local administration of ADSCs after cuff repair. Additional preclinical and clinical studies are necessary to elucidate the optimal utilization of progenitor cells to enhance tendon healing.

Scaffolds

An area of recent interest in research is the development of new ways to produce synthetic, degradable scaffolds that



TABLE I Grades of Recommendations for Clinical Care*†						
	Microfracture	PRP	ACI/ MACI	Progenitor Cells	Bone- Marrow Venting	Scaffolds
Focal chondral defects	Grade B	Grade I	Grade I	Grade I	Grade I	NA
Diffuse osteoarthritis	NA	Grade C	NA	Grade I	NA	NA
Rotator cuff repair	Grade C	Grade C	NA	Grade I	Grade I	Grade I

*Grade A indicates good evidence (Level-I studies with consistent findings) for or against recommending intervention. Grade B indicates fair evidence (Level-II or III studies with consistent findings) for or against recommending intervention. Grade C indicates conflicting or poor-quality evidence (Level-IV or V studies) not allowing a recommendation for or against intervention. Grade I indicates that there is insufficient evidence to make a recommendation. †PRP = platelet-rich plasma, ACI = autologous chondrocyte implantation, MACI = matrix-induced autologous chondrocyte implantation, and NA = not applicable.

reproduce the function and structure of the rotator cuff tendon. Notable advances include the recent development of 3-dimensional electrospun scaffolds that closely resemble extracellular matrix. In addition, coating and fabrication techniques that facilitate the integration of bioactive molecules, including growth factors, within scaffolds have been developed^{57,58}. Although these scaffolds are not ready for human implantation at this time, collaboration between researchers focusing on cell biology, biomaterial science, and tissue engineering may lead to scaffolds with the requisite characteristics to initiate rotator cuff tendon regeneration. Other recently developed types of scaffold for the augmented repair of massive rotator cuff tears or the replacement of irreparable cuff tissue are human acellular dermal allograft^{59,60} and xenograft⁶¹. In addition to biomechanical strengthening of the tendon tissue, the rationale behind these techniques is to render the graft acellular, decreasing its immunogenicity, while leaving an intact collagen extracellular matrix; the intact matrix thereby promotes growth of new host tissue into the graft⁶². Short-term results after 24 months of follow-up have been promising^{59,61-63}; Barber et al., for example, in a randomized prospective trial, demonstrated superior outcome scores and a lower failure rate with augmented compared with non-augmented rotator cuff repairs⁶². However, further studies are necessary to evaluate the longterm benefits of human acellular dermal scaffolds.

Conclusions

In summary, basic-science studies investigating biological factors to treat chondral defects or enhance rotator cuff healing show promising results. However, clinical data are still limited by inconsistency and controversy, and recommendations for clinical care are not possible (Table I). For focal chondral lesions of the glenoid or humeral head, microfracture has been reported to yield positive postoperative results in the intermediate to long-term follow-up period. The use of PRP, ACI, and MACI treatments for shoulder chondral lesions also has been reported; however, no conclusions can be drawn concerning the efficacy of these modalities. For rotator cuff tear healing, animal and basicscience studies have demonstrated that modulation of growth factors and progenitor cells may be helpful, but the optimal modulation and delivery vehicle remain to be elucidated. Conflicting results have been reported about the role of PRP as biological augmentation for rotator cuff repairs, making this subject an area of continuing debate. In general, additional clinical trials are necessary to elucidate the ideal biological approaches to improve the healing of a variety of musculoskeletal tissues.

Jonas Pogorzelski, MD, MHBA¹, Jonathan A. Godin, MD, MBA^{1,2}, Erik M. Fritz, MD¹, Mark E. Cinque, MS¹, Jorge Chahla, MD¹, Johnny Huard, PhD^{1,3}, Peter J. Millett, MD, MSc^{1,2} ¹Steadman Philippon Research Institute, Vail, Colorado

²The Steadman Clinic, Vail, Colorado

³Department of Orthopaedic Surgery, McGovern School of Medicine, University of Texas Health Science Center at Houston, Houston, Texas

E-mail address for P.J. Millett: drmillett@thesteadmanclinic.com

References

- **1.** Pope DP, Croft PR, Pritchard CM, Silman AJ. Prevalence of shoulder pain in the community: the influence of case definition. Ann Rheum Dis. 1997 May;56(5):308-12.
- Millett PJ, Huffard BH, Horan MP, Hawkins RJ, Steadman JR. Outcomes of full-thickness articular cartilage injuries of the shoulder treated with microfracture. Arthroscopy. 2009 Aug;25(8):856-63.
- **3.** Wang VM, Karas V, Lee AS, Yin Z, Van Thiel GS, Hussey K, Sumner DR, Chubinskaya S, Magin RL, Verma NN, Romeo AA, Cole BJ. Assessment of glenoid chondral healing: comparison of microfracture to autologous matrix-induced chondrogenesis in a novel rabbit shoulder model. J Shoulder Elbow Surg. 2015 Nov;24(11): 1789-800. Epub 2015 Aug 1.
- 4. Steadman JR, Briggs KK, Matheny LM, Guillet A, Hanson CM, Willimon SC. Outcomes following microfracture of full-thickness articular cartilage lesions of the knee in adolescent patients. J Knee Surg. 2015 Apr;28 (2):145-50. Epub 2014 Apr 24.
- 5. Steadman JR, Hanson CM, Briggs KK, Matheny LM, James EW, Guillet A. Outcomes after knee microfracture of chondral defects in alpine ski racers. J Knee Surg. 2014 Oct;27(5): 407-10. Epub 2014 May 22.
- **6.** Steadman JR, Matheny LM, Briggs KK, Rodkey WG, Carreira DS. Outcomes following healing response in older, active patients: a primary anterior cruciate ligament repair technique. J Knee Surg. 2012 Jul;25(3):255-60.
- 7. Steadman JR, Rodkey WG, Rodrigo JJ. Microfracture: surgical technique and rehabilitation to treat chondral defects. Clin Orthop Relat Res. 2001 Oct;(391)(Suppl):S362-9.



- 8. Goyal D, Keyhani S, Lee EH, Hui JH. Evidencebased status of microfracture technique: a systematic review of level I and II studies. Arthroscopy. 2013 Sep;29(9):1579-88.
- Richards RR, An KN, Bigliani LU, Friedman RJ, Gartsman GM, Gristina AG, Iannotti JP, Mow VC, Sidles JA, Zuckerman JD. A standardized method for the assessment of shoulder function. J Shoulder Elbow Surg. 1994 Nov;3(6): 347-52. Epub 2009 Feb 13.
- 10. Frank RM, Van Thiel GS, Slabaugh MA, Romeo AA, Cole BJ, Verma NN. Clinical outcomes after microfracture of the glenohumeral joint. Am J Sports Med. 2010 Apr; 38(4):772-81. Eoub 2010 Jan 21.
- 11. Siebold R, Lichtenberg S, Habermeyer P. Combination of microfracture and periostal-flap for the treatment of focal full thickness articular cartilage lesions of the shoulder: a prospective study. Knee Surg Sports Traumatol Arthrosc. 2003 May;11(3):183-9. Epub 2003 Apr 29
- **12.** Greenspoon JA, Moulton SG, Millett PJ, Petri M. The role of platelet rich plasma (PRP) and other biologics for rotator cuff repair. Open Orthop J. 2016 Jul 21;10:309-14.
- 13. Warth RJ, Dornan GJ, James EW, Horan MP, Millett PJ. Clinical and structural outcomes after arthroscopic repair of full-thickness rotator cuff tears with and without platelet-rich product supplementation: a meta-analysis and meta-regression. Arthroscopy. 2015 Feb;31(2): 306-20. Epub 2014 Nov 14.
- 14. Halpern B, Chaudhury S, Rodeo SA, Hayter C, Bogner E, Potter HG, Nguyen J. Clinical and MRI outcomes after platelet-rich plasma treatment for knee osteoarthritis. Clin J Sport Med. 2013 May;23(3):238-9.
- **15.** Metcalf KB, Mandelbaum BR, McIlwraith CW. Application of platelet-rich plasma to disorders of the knee joint. Cartilage. 2013 Oct;4 (4):295-312.
- **16.** LaPrade RF, Geeslin AG, Murray IR, Musahl V, Zlotnicki JP, Petrigliano F, Mann BJ. Biologic treatments for sports injuries II think tank-current concepts, future research, and barriers to advancement, part 1: biologics overview, ligament injury, tendinopathy. Am J Sports Med. 2016 Dec;44(12):3270-83. Epub 2016
- 17. Murray IR, LaPrade RF, Musahl V, Geeslin AG, Zlotnicki JP, Mann BJ, Petrigliano FA. Biologic treatments for sports injuries II think tank-current concepts, future research, and barriers to advancement, part 2: rotator cuff. Orthop J Sports Med. 2016 Mar 31;4(3): 2325967116636586.
- **18.** Zavadil DP, Satterlee CC, Costigan JM, Holt DW, Shostrom VK. Autologous platelet gel and platelet-poor plasma reduce pain with total shoulder arthroplasty. J Extra Corpor Technol. 2007 Sep;39(3):177-82.
- **19.** Lo EY, Flanagin BA, Burkhead WZ. Biologic resurfacing arthroplasty with acellular human dermal allograft and platelet-rich plasma (PRP) in young patients with glenohumeral arthritisaverage of 60 months of at mid-term follow-up. J Shoulder Elbow Surg. 2016 Jul;25(7):e199-207. Epub 2016 Feb 17.
- **20.** Basad E, Wissing FR, Fehrenbach P, Rickert M, Steinmeyer J, Ishaque B. Matrix-induced autologous chondrocyte implantation (MACI) in the knee: clinical outcomes and challenges. Knee Surg Sports Traumatol Arthrosc. 2015 Dec; 23(12):3729-35. Epub 2014 Sep 14.

- **21.** Romeo AA, Cole BJ, Mazzocca AD, Fox JA, Freeman KB, Joy E. Autologous chondrocyte repair of an articular defect in the humeral head. Arthroscopy. 2002 Oct;18(8):925-9.
- **22.** Buchmann S, Salzmann GM, Glanzmann MC, Wörtler K, Vogt S, Imhoff AB. Early clinical and structural results after autologous chodrocyte transplantation at the glenohumeral joint. J Shoulder Elbow Surg. 2012 Sep;21(9):1213-21. Epub 2011 Nov 2.
- **23.** Jacobi M, Villa V, Magnussen RA, Neyret P. MACI a new era? Sports Med Arthrosc Rehabil Ther Technol. 2011 May 20;3(1):10-10.
- 24. Lorbach O, Baums MH, Kostuj T, Pauly S, Scheibel M, Carr A, Zargar N, Saccomanno MF, Milano G. Advances in biology and mechanics of rotator cuff repair. Knee Surg Sports Traumatol Arthrosc. 2015 Feb;23(2):530-41. Epub 2015 Jan 9.
- **25.** Snyder SJ, Burns J. Rotator cuff healing and the bone marrow "crimson duvet" from clinical observations to science. Tech Shoulder Elbow Surg. 2009;10:130-7.
- **26.** Gotoh M, Mitsui Y, Shibata H, Yamada T, Shirachi I, Nakama K, Okawa T, Higuchi F, Nagata K. Increased matrix metalloprotease-3 gene expression in ruptured rotator cuff tendons is associated with postoperative tendon retear. Knee Surg Sports Traumatol Arthrosc. 2013 Aug;21(8):1807-12. Epub 2012 Sep 22.
- 27. Ide J, Kikukawa K, Hirose J, Iyama K, Sakamoto H, Fujimoto T, Mizuta H. The effect of a local application of fibroblast growth factor-2 on tendon-to-bone remodeling in rats with acute injury and repair of the supraspinatus tendon. J Shoulder Elbow Surg. 2009 May-Jun; 18(3):391-8.
- **28.** Uggen C, Dines J, McGarry M, Grande D, Lee T, Limpisvasti O. The effect of recombinant human platelet-derived growth factor BB-coated sutures on rotator cuff healing in a sheep model. Arthroscopy. 2010 Nov;26(11): 1456-62. Epub 2010 Aug 21.
- 29. Bedi A, Fox AJ, Kovacevic D, Deng XH, Warren RF, Rodeo SA. Doxycycline-mediated inhibition of matrix metalloproteinases improves healing after rotator cuff repair. Am J Sports Med. 2010 Feb;38(2):308-17. Epub 2009 Oct 13.
- **30.** Osti L, Del Buono A, Maffulli N. Microfractures at the rotator cuff footprint: a randomised controlled study. Int Orthop. 2013 Nov;37(11):2165-71. Epub 2013 Jun 13.
- **31.** Milano G, Saccomanno MF, Careri S, Taccardo G, De Vitis R, Fabbriciani C. Efficacy of marrow-stimulating technique in arthroscopic rotator cuff repair: a prospective randomized study. Arthroscopy. 2013 May;29(5):802-10. Epub 2013 Mar 21.
- **32.** Jo CH, Kim JE, Yoon KS, Shin S. Platelet-rich plasma stimulates cell proliferation and enhances matrix gene expression and synthesis in tenocytes from human rotator cuff tendons with degenerative tears. Am J Sports Med. 2012 May;40(5):1035-45. Epub 2012 Feb 23.
- **33.** Sadoghi P, Lohberger B, Aigner B, Kaltenegger H, Friesenbichler J, Wolf M, Sununu T, Leithner A, Vavken P. Effect of platelet-rich plasma on the biologic activity of the human rotator-cuff fibroblasts: a controlled in vitro study. J Orthop Res. 2013 Aug;31 (8):1249-53. Epub 2013 Apr 8.
- **34.** Namazi H. Rotator cuff repair healing influenced by platelet-rich plasma construct

- augmentation: a novel molecular mechanism. Arthroscopy. 2011 Nov;27(11):1456, author reply:1456-7.
- **35.** Antuña S, Barco R, Martínez Diez JM, Sánchez Márquez JM. Platelet-rich fibrin in arthroscopic repair of massive rotator cuff tears: a prospective randomized pilot clinical trial. Acta Orthop Belg. 2013 Feb;79(1):25-30.
- 36. Randelli P, Arrigoni P, Ragone V, Aliprandi A, Cabitza P. Platelet rich plasma in arthroscopic rotator cuff repair: a prospective RCT study, 2-year follow-up. J Shoulder Elbow Surg. 2011 Jun;20(4):518-28.
- **37.** Rodeo SA, Delos D, Williams RJ, Adler RS, Pearle A, Warren RF. The effect of platelet-rich fibrin matrix on rotator cuff tendon healing: a prospective, randomized clinical study. Am J Sports Med. 2012 Jun;40(6):1234-41. Epub 2012 Apr 10.
- **38.** Ruiz-Moneo P, Molano-Muñoz J, Prieto E, Algorta J. Plasma rich in growth factors in arthroscopic rotator cuff repair: a randomized, double-blind, controlled clinical trial. Arthroscopy. 2013 Jan;29(1):2-9.
- **39.** Malavolta EA, Gracitelli ME, Ferreira Neto AA, Assunção JH, Bordalo-Rodrigues M, de Camargo OP. Platelet-rich plasma in rotator cuff repair: a prospective randomized study. Am J Sports Med. 2014 Oct;42(10):2446-54. Epub 2014 Aug 1.
- **40.** Charousset C, Zaoui A, Bellaïche L, Piterman M. Does autologous leukocyte-platelet-rich plasma improve tendon healing in arthroscopic repair of large or massive rotator cuff tears? Arthroscopy. 2014 Apr;30(4):428-35.
- **41.** Wang A, McCann P, Colliver J, Koh E, Ackland T, Joss B, Zheng M, Breidahl B. Do postoperative platelet-rich plasma injections accelerate early tendon healing and functional recovery after arthroscopic supraspinatus repair? A randomized controlled trial. Am J Sports Med. 2015 Jun;43(6):1430-7. Epub 2015 Mar 19
- **42.** Gwinner C, Gerhardt C, Haneveld H, Scheibel M. Two-staged application of PRP in arthroscopic rotator cuff repair: a matched-pair analysis. Arch Orthop Trauma Surg. 2016 Aug; 136(8):1165-71. Epub 2016 Jul 5.
- 43. Flury M, Rickenbacher D, Schwyzer HK, Jung C, Schneider MM, Stahnke K, Goldhahn J, Audigé L. Does pure platelet-rich plasma affect postoperative clinical outcomes after arthroscopic rotator cuff repair? A randomized controlled trial. Am J Sports Med. 2016 Aug;44 (8):2136-46. Epub 2016 May 16.
- **44.** Pandey V, Bandi A, Madi S, Agarwal L, Acharya KK, Maddukuri S, Sambhaji C, Willems WJ. Does application of moderately concentrated platelet-rich plasma improve clinical and structural outcome after arthroscopic repair of medium-sized to large rotator cuff tear? A randomized controlled trial. J Shoulder Elbow Surg. 2016 Aug;25(8):1312-22. Epub 2016 Jun 1.
- **45.** Jo CH, Shin JS, Shin WH, Lee SY, Yoon KS, Shin S. Platelet-rich plasma for arthroscopic repair of medium to large rotator cuff tears: a randomized controlled trial. Am J Sports Med. 2015 Sep;43(9):2102-10. Epub 2015 May 26.
- **46.** Holtby R, Christakis M, Maman E, MacDermid JC, Dwyer T, Athwal GS, Faber K, Theodoropoulos J, Woodhouse LJ, Razmjou H. Impact of platelet-rich plasma on arthroscopic repair of small- to medium-sized rotator cuff tears: a randomized controlled trial. Orthop J



- Sports Med. 2016 Sep 13;4(9): 2325967116665595.
- **47.** Vavken P, Sadoghi P, Palmer M, Rosso C, Mueller AM, Szoelloesy G, Valderrabano V. Platelet-rich plasma reduces retear rates after arthroscopic repair of small- and medium-sized rotator cuff tears but is not cost-effective. Am J Sports Med. 2015 Dec;43(12):3071-6. Epub 2015 Mar 12.
- **48.** Chahal J, Van Thiel GS, Mall N, Heard W, Bach BR, Cole BJ, Nicholson GP, Verma NN, Whelan DB, Romeo AA. The role of platelet-rich plasma in arthroscopic rotator cuff repair: a systematic review with quantitative synthesis. Arthroscopy. 2012 Nov;28(11):1718-27. Epub 2012 Jun 12
- **49.** Saltzman BM, Jain A, Campbell KA, Mascarenhas R, Romeo AA, Verma NN, Cole BJ. Does the use of platelet-rich plasma at the time of surgery improve clinical outcomes in arthroscopic rotator cuff repair when compared with control cohorts? A systematic review of meta-analyses. Arthroscopy. 2016 May;32(5): 906-18. Epub 2015 Dec 23.
- **50.** Lhee SH, Jo YH, Kim BY, Nam BM, Nemeno JG, Lee S, Yang W, Lee Jl. Novel supplier of mesenchymal stem cell: subacromial bursa. Transplant Proc. 2013 Oct;45(8):3118-21.
- **51.** Song N, Armstrong AD, Li F, Ouyang H, Niyibizi C. Multipotent mesenchymal stem cells from human subacromial bursa: potential for cell based tendon tissue engineering. Tissue Eng Part A. 2014 Jan; 20(1-2):239-49. Epub 2013 Aug 21.

- **52.** Gulotta LV, Kovacevic D, Ehteshami JR, Dagher E, Packer JD, Rodeo SA. Application of bone marrow-derived mesenchymal stem cells in a rotator cuff repair model. Am J Sports Med. 2009 Nov;37(11):2126-33. Epub 2009 Aug 14.
- **53.** Kida Y, Morihara T, Matsuda K, Kajikawa Y, Tachiiri H, Iwata Y, Sawamura K, Yoshida A, Oshima Y, Ikeda T, Fujiwara H, Kawata M, Kubo T. Bone marrow-derived cells from the footprint infiltrate into the repaired rotator cuff. J Shoulder Elbow Surg. 2013 Feb;22(2):197-205. Epub 2012 Apr 28.
- **54.** Oh JH, Chung SW, Kim SH, Chung JY, Kim JY. 2013 Neer Award: effect of the adipose-derived stem cell for the improvement of fatty degeneration and rotator cuff healing in rabbit model. J Shoulder Elbow Surg. 2014 Apr;23(4): 445-55. Epub 2013 Oct 12.
- **55.** Ellera Gomes JL, da Silva RC, Silla LM, Abreu MR, Pellanda R. Conventional rotator cuff repair complemented by the aid of mononuclear autologous stem cells. Knee Surg Sports Traumatol Arthrosc. 2012 Feb;20(2):373-7. Epub 2011 Jul 20.
- **56.** Hernigou P, Flouzat Lachaniette CH, Delambre J, Zilber S, Duffiet P, Chevallier N, Rouard H. Biologic augmentation of rotator cuff repair with mesenchymal stem cells during arthroscopy improves healing and prevents further tears: a case-controlled study. Int Orthop. 2014 Sep;38(9):1811-8. Epub 2014 Jun 7.
- **57.** Breidenbach AP, Gilday SD, Lalley AL, Dyment NA, Gooch C, Shearn JT, Butler DL. Functional tissue engineering of tendon: establishing biological success criteria for

- improving tendon repair. J Biomech. 2014 Jun 27;47(9):1941-8. Epub 2013 Oct 22.
- **58.** Hakimi O, Murphy R, Stachewicz U, Hislop S, Carr AJ. An electrospun polydioxanone patch for the localisation of biological therapies during tendon repair. Eur Cell Mater. 2012 Oct 23;24:344-57, discussion :357.
- 59. Petri M, Warth RJ, Horan MP, Greenspoon JA, Millett PJ. Outcomes after open revision repair of massive rotator cuff tears with biologic patch augmentation. Arthroscopy. 2016 Sep;32 (9):1752-60. Epub 2016 Apr 6.
- **60.** Petri M, Greenspoon JA, Moulton SG, Millett PJ. Patch-augmented rotator cuff repair and superior capsule reconstruction. Open Orthop J. 2016 Jul 21;10:315-23.
- **61.** Bokor DJ, Sonnabend D, Deady L, Cass B, Young A, Van Kampen C, Arnoczky S. Preliminary investigation of a biological augmentation of rotator cuff repairs using a collagen implant: a 2-year MRI follow-up. Muscles Ligaments Tendons J. 2015 Oct 20;5(3): 144-50
- **62.** Barber FA, Burns JP, Deutsch A, Labbé MR, Litchfield RB. A prospective, randomized evaluation of acellular human dermal matrix augmentation for arthroscopic rotator cuff repair. Arthroscopy. 2012 Jan;28(1):8-15. Epub 2011 Oct 5.
- **63.** Greenspoon JA, Millett PJ, Moulton SG, Petri M. Irreparable rotator cuff tears: restoring joint kinematics by tendon transfers. Open Orthop J. 2016 Jul 21;10:266-76.